

APPLICATIONS OF ULTRA HIGH PRESSURE LIQUID CHROMATOGRAPHY TO NATURAL PRODUCTS

Shradhanjali Singh*, Anil Kumar Singh¹

*Department of Pharmaceutical Chemistry, United Institute of Pharmacy, A-31/1 UPSIDC, Industrial Area, Naini Allahabad, Uttar Pradesh-211010

¹ Department of pharmaceuticals, United Institute of Pharmacy, A-31/1 UPSIDC, Industrial Area, Naini Allahabad, Uttar Pradesh-211010

ABSTRACT

Nature has afforded a number of interesting chemicals to be used in medicine, cosmetics, agriculture and others. The success in the studies of complex extracts of natural products depends on the development of new analytical techniques for the analysis of extracts. In view of increased separation efficiency, improved resolution and faster analysis time, Liquid chromatography established a method involving smaller particle size (<2µm) and high pressure (> 350bar). With the introduction of Ultra-High Pressure Liquid Chromatography (UHPLC), in 2004 new opportunities are emerging in the pharmaceutical industry for obtaining rapid analytical separations without sacrificing high-quality results in terms of resolution, accuracy, and reliability. To ensure the accurate quantification of the selected marker compound in natural products hyphenated techniques includes UHPLC-MS, SFE-UHPLC and UHPLC-DAD-ESI-MS. The present review seeks a newly developed analytical method as Ultra High Pressure Liquid chromatography together with the hyphenated techniques and its applications to natural products. This technique is very versatile and powerful tool for the separation of natural product from crude extracts for selective detection and general profiling. The technique is precise, robust, faster and sensitive and relies upon smaller volumes of organic solvents than HPLC. The advantages of introducing UHPLC are a decrease in sample turnaround time for both manufacturing and product development, the use of less organic solvents, and a reduction in generated waste.

Key words: UHPLC, natural product, hyphenated techniques, product development.

INTRODUCTION

Plants have always a rich source of active compounds which needs to be isolated for the activity. Natural products contain the active compounds which are complex structures and need to be separated. The separation can be achieved by using a well diversified analytical technique. Several analytical techniques have been employed for the separation of natural compounds like Spectrophotometric methods, Gas chromatography, Supercritical fluid chromatography, Capillary electrophoresis, High performance liquid chromatography, Ultra High Pressure liquid Chromatography. Ultra High Pressure liquid Chromatography is a developing technique based on the principle of separating the compounds based on increasing the resolving power of the analytical separation process particularly with the development of columns packed with porous sub 2micron meter particles used in very high pressure conditions. In this technique the use of smaller particle size results in high plate numbers as well as faster separations. This can be shown in Equation-1¹

$$H = A + \frac{B}{\mu} + C\mu \quad (1)$$

Since in the last eight years UHPLC technique is increasing widely for the separation as compared to HPLC technique because it eliminates complications resulting from use of larger samples volume, larger particle size, increased generated wastes and increased analysis time². By comparing the intrinsic performance of such packing size with other intrinsic techniques such as monoliths, fused core technology or high temperature liquid chromatography with conventional particle size UHPLC with a maximum pressure of 1000bar is a very attractive strategy that

***Corresponding author:**

Email: singhshradhanjali@gmail.com

generates a lowest analysis time for the separation. HPLC method is developed and validated for those plant product which are previously known and do not having co-eluting analytes but for newer plants extracts, there is need to develop a method which is having a greater resolution and lesser analysis time³. This review describes the use of Ultra High Pressure Liquid Chromatography (UHPLC) in analyzing natural products.

Since last five year's number of publications on the theory, principles, techniques and applications of UHPLC have been reported. UHPLC technique is considered as a valuable technique for the separation of natural products and can be coupled with various types of detectors for the various quantitative and qualitative analyses of natural products⁴. The application of this technique is in the field of herbal medicine and dietary supplement⁴, food technology⁵, explosive compounds⁶, cosmetic products have been proved. By comparing its intrinsic performance with other existing techniques such as monoliths or high temperature liquid chromatography, it is very attractive strategy for improving chromatographic efficiency in the range 1000-80,000 plates^{7, 8}. The tubing volumes, tubing length, gradient delay volume, column dimensions, mobile phase composition, injection volume, and detector cell volume are the parameters which are needed to be controlled for efficient separation. Improvements on these conditions have become the major themes in the development of UHPLC techniques.

Numerous studies for dealing with these problems have been published recently and many applications have been used to illustrate and validate the feasibility of the use of UHPLC technique.

A column with perfluorophenyl functionality helps to enhance retention and improve resolution when separating complex mixtures of substituted aromatic compounds⁹. The concept of making the column stable by using such a high pressure and the sample loading problem was solved using ethyl bridged hybrid particles for UHPLC column packing¹⁰. Analysis of substances with a wide range of polarities in a single run is estimated with ultra high pressure liquid chromatography composed of capillary packed column. The columns were packed with 1.5 μ m nonporous octadecyl silane modified silica particles¹¹. To analyse and establish the impurity profile in the natural product, to decrease

extracolumn band dispersion, to increase the mass loadability the smaller internal diameter of a column in mm is needed for proper separation¹². A major application of this technique is in the estimation of natural compounds in traditional Chinese medicine (TCM). TCM has many complex structures containing natural compounds and metabolomics which are to be separated and for separation of these compounds a profound detector is must. Various hyphenated techniques is being developed are UPLC/MS-MS¹³⁻¹⁵, UPLC-PDA¹⁶, UPLC- TOF-MS¹⁷.

Apart from the separation method described above there are other methods for separation also, Liquid extraction method to separate flavonoids from plant extract using UHPLC method¹⁸ proves to be significant among the separation. Separation of some compounds requires LC-MS method in conjunction with fast atom bombardment¹⁹, thermospray²⁰ and atmospheric-pressure ionization technique such as electrospray ionization²¹⁻³⁶ to avoid degradation due to GC-MS problems. Many papers have been published regarding the separation of proteins viz, High performance liquid chromatographic method, Ion exchange chromatography but the use of UHPLC method using monoliths column is also widespread in the field of separation of proteins by using UV detector³⁷. For complex extracts different elution methods can be used to improve the resolution power significantly. These are classified as isocratic system, gradient system³⁸. Simple UHPLC-UV³⁹ method, a UHPLC-TOF LC/MS⁴⁰ method for degradative samples of natural products has also used as a wide application in the separation of natural product. The content determination and fingerprinting analysis of TCMS can be performed efficiently by UHPLC technique for various active compounds⁴¹. Various techniques used in pre and post column is often used to overcome the separation and detection of compounds⁴².

NATURAL PRODUCT SEPARATION BY UHPLC METHOD

Gradient elution system is widely used in the separation of non polar as well as polar substituents in a compound. Separation of phenolic antioxidants and ascorbyl palmitate from edible oils is done on two columns. The sample of edible oils was eluted from the HPLC column C18 100 x 4.6 mm, 5 μ m and analytical C18, 1.9 μ m 50 mm x 2.1 mm column using a gradient system having Mobile phase A 70:30v/v, Acetonitrile and methanol and mobile phase B 1% phosphoric acid in water for HPLC

column and Mobile phase A 70:30v/v, Acetonitrile and methanol and mobile phase B 0.02% formic acid in water for UHPLC column. It was observed that 70% reduction in run time and 90% reduction in solvent usage were there by using UHPLC column as compared to HPLC column⁴³. The separation is shown in Figure 1. By Liquid chromatography the profiling of crude extracts from plant origin or from other biological sources are easy to evolve into powerful tools for dereplication, quality assessment and metabolomics. Metabolite profiling of crude extracts can be easily done by high performance

neutraceutical by UHPLC method is done by using 1.5µm particle 50 mm length C18 columns and at UV-Visible detector at 254nm⁴⁵.

The traditional Spectroscopic method for the screening of Xanthine oxidase inhibitor and superoxide anion scavenger is been widely used but using UHPLC-TQ-MS the estimation can be done in single analysis⁴⁶. Detailed metabolite profiling and dereplication of crude plant extracts is mandatory for both Quality control and metabolomics purpose requires high resolution separation and sensitive detection and therefore to do so UHPLC-TOF MS

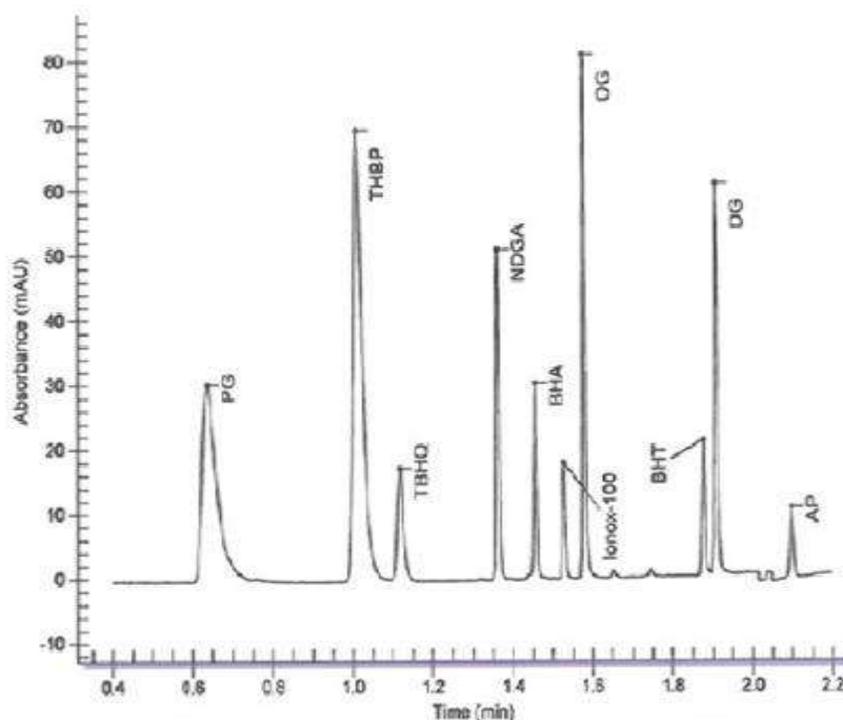


Figure 1 Separation of 10 antioxidants in edible oils viz., PG (propyl gallate), THBP (2, 4, 5-trihydroxybutyrophenone), TBHQ (t-butylhydroquinone), NDGA (nordihydroguaric acid), BHA (2 (or 3)-tert-butyl-4-hydroxyanisole), OG (Octyl gallate), BHT (butylated hydroxy toluene), DG (dodecyl gallate), AP (Ascorbyl palmitate) by UHPLC method.

liquid chromatography (HPLC), such as liquid chromatography photodiode array detection (LC-PDA), mass spectrometry (LC-MS) or nuclear magnetic resonance (LC-NMR)⁴⁴.

The determination of neutraceuticals in natural compound by HPC method had a disadvantage of using larger run time and larger volume of mobile phase. This problem can be overcome by using UHPLC method. Determination of isoflavones in

detector was used at higher temperature. The percentage reduction was observed less as compared to the previous methods⁴⁷. Conventional HPLC method provides insufficient resolving power for phenolics compounds due to the complex nature of natural compounds. So to overcome this process a new advanced UHPLC method was developed and for the phenolic compound⁴⁸.

The development of high resolution methods related to HPLC for both chemical and biological profile has significantly increased the efficiency of classical bioactivity guided fractional procedures. A UHPLC method has been developed for the natural product drug discovery for nutraceuticals⁴⁹.

CONCLUSION

Ultra high performance chromatography has found its use for the separation of components found in natural products. The fact that UHPLC uses smaller particle size has decreased the analysis time for the separation is a big advantage that encompasses the problems of band broadening. Considering the savings of mobile phases the separation technique by using UHPLC method is being widely used in the future era of natural product evaluation.

REFERENCES

1. Mellors, J.C and J.W.Jorgenson, 2004. Use of 1.5µm porous ethyl-bridged hybrid particles as a stationary phase support for Reversed phase Ultra high pressure liquid chromatography. *Anal.Chem*, 76: 5441-5450.
2. Hill, Lionel and T.I. Wang., 2009. Approaches to the analysis of plant derived natural products. In *plant derived natural products, synthesis, functions and application*, Eds., Osbourn, A.E and V, Lanzotti, Springer science: Business media New York publishers, pp: 104-105.
3. Davy, G and J.L. Veuthoy, 2011, Guidelines for the use of UHPLC instruments, laboratory of analytical pharmaceutical chemistry, school of pharmaceutical sciences, University of Geneva, Switzerland.
4. G, Du., H.Y. Zhao., Q.W. Zhang., G.H. Li.,Y.C. Li and X.Y. Wang, 2010. A rapid method for Simultaneous determination of 14 phenolic compounds in radix Puerariae using microwave assisted extraction and UHPLC coupled with diode array detection and time of flight mass spectrometry. *Journal of chromatography A*, 1217: 705-714.
5. Njies, P, 2011, Rapid UHPLC determination of common antioxidants in edible oils, Shelton CT 06484 USA.
6. Guifeng, J, 2012, Simultaneous UHPLC/MS analyses of explosive compounds, Thermofischer scientific, San jose, CA, USA, 51879.
7. Dolan, J.W., L.R. Snyder, 1998. Maintaining fixed band spacing when changing column dimensions in gradient elution. *Journal of chromatography A*, 799: 21-34.
8. Schellinger, A.P and P.W. Carr, 2005. A practical approach to transferring linear gradient elution methods, *Journal of chromatography A*, 1077: 110-119.
9. Guifeng, J and J. Zhang, 2010, Quantitative analysis of catechins in tea by UHPLC/UV, Thermofischer scientific, San jose, CA, USA, 52038.
10. Mellors, J.C and J.W. Jorgenson, 2004. Use of 1.5µm porous ethyl bridged hybrid particles as a stationary phase support for RP-UPLC, *Anal.Chem*, 76: 5441-5450.
11. Macnair, J.E., L.C. Lewis and J.W. Jorgenson, 1997. Ultra High pressure reversed phase liquid chromatography in packed capillary column, *Anal.Chem*, 69: 983-989.
12. Colon, L.A., J.M. Cintron, J.A.Anspach, A.M. Fermier and A.S. Kelly, 2004. Very high pressure HPLC with 1 mm id columns, *Analyst*, 129: 503-504.
13. Frenich, A.G., J.L.V. Martinez, R.R. Gonzalez, 2008. Simple and high throughput method for the multimycotoxin analysis in cereals and related foods by UHPLC/ MS, *Food Chemistry*, 117(4): 705-712.
14. Gardana, C., M.Scaglianti, P. Simoneth, 2010. Evaluation of steviol and its glycosides in stevia rubaudiana leaves and commercial sweetener by UHPLC-MS, *Journal of chromatography A*, 1217(3): 1463-1470.
15. Ceymann, M., E. Arrigoni, H. scharer, D. Baumgartner, R.F. Hurrell, 2011. Rapid high performance Screening method using UHPLC-MS to quantify 12 polyphenol compounds in fresh apples, *Analytical methods*, 3: 1774-1778.
16. Chen, J., F. wang, J. Liu, F. Sen-chun Lee, X. Wang, H.Yang, 2008. Analysis of alkaloids in coptis ehinensis franch by accelerated solvent extraction combined with UHPLC analysis with PDA and Tandem MS detectors, *Analytica Chemica Acta*, 613(2): 184-195.
17. Chan, E., et.al, 2007. Ultra high pressure liquid chromatography / TOF-MS based metabolomics of raw steamed panax notoginseng, *Rapid communication in MS*, 21(4): 519-528.
18. Chen, X.J., H. Ji, Q.W.Zhang, P.F.Tu, Y.T. Wang, 2008. A rapid method for simultaneous determination of 15 flavonoids in epimedium using pressurized liquid extraction and UPLC, *Journal of pharmaceutical and biomedical analysis*, 46(2): 226-235.

19. Yeung, P., P. Vouros, GS. Reddy, 1993. Characterization of vitamin D₃ metabolites using continuous-flow fast atom bombardment tandem mass spectrometry and high-performance liquid chromatography, *J Chromatogr*, 645: 115–123.
20. Watson, D., KD. Setchell, R, Ross, 1991. Analysis of vitamin D and its metabolites using thermospray liquid chromatography/mass spectrometry, *Biomed Chromatogr*, 5: 153–160.
21. Kushnir, MM., JA.Ray, AL.Rockwood, WL. Roberts, SL. La’ulu, JE. Whittington, AW. Meike, 2010. Rapid analysis of 25-hydroxyvitamin D(2) and D(3) by liquid chromatography-tandem mass spectrometry and association of vitamin D and parathyroid hormone concentrations in healthy adults, *Am J Clin Pathol*, 134: 148–156.
22. Ouweland, JM., AM. Beijers, PN. Demacker, H. Daal, 2010. Measurement of 25-OH-vitamin D in human serum using liquid chromatography tandem-mass spectrometry with comparison to radioimmunoassay and automated immunoassay, *J Chromatogr B*, 878: 1163–1168.
23. Vogeser, M. 2010. *J Steroid Biochem Mol Biol* (in press).
24. Duan, X., B. Weinstock-Guttman, H.Wang, E. Bang, R. Li, M. Ramanathan, J. Qu, 2010. Ultrasensitive quantification of serum vitamin D metabolites using selective solid-phase extraction coupled to microflow liquid chromatography and isotope-dilution mass spectrometry, *Anal Chem*, 82: 2488–2497.
25. Tai, S.S., M. Bedner, K.W. Phinney, 2010. Development of a Candidate Reference Measurement Procedure for the Determination of 25-hydroxyvitamin D₃ and 25-hydroxy vitamin D₂ in Human Serum Using Isotope-Dilution Liquid Chromatography / Tandem Mass Spectrometry, *Anal Chem*, 82: 1942–1948.
26. Singh, R.J, 2010. Quantitation of 25-OH-vitamin D (25OHD) using liquid tandem mass spectrometry (LC-MS-MS), *Methods Mol Biol*, 603: 509–517.
27. Hojskov, C.S., L. Heickendorf, H.J. Moller, 2010. High-throughput liquid-liquid extraction and LCMSMS assay for determination of circulating 25(OH) vitamin D₃ and D₂ in the routine clinical laboratory, *Clin Chim Acta*, 411: 114–116.
28. Bunch, D.R., A.Y. Miller, S. Wang, 2009. Development and validation of a liquid chromatography-tandem mass spectrometry assay for serum 25-hydroxyvitamin D₂/D₃ using a turbulent flow online extraction technology, *Clin Chem Lab Med*, 47: 1565–1572.
29. Knox, S., J. Harris, L. Carlton, A.M. Wallace, 2009. A simple automated solid-phase extraction procedure for measurement of 25-hydroxyvitamin D₃ and D₂ by liquid chromatography-tandem mass spectrometry, *Ann Clin Biochem*, 46: 226–230.
30. Roth, H.J., H. Schmidt-Gayk, H. Weber, C. Niedrau, 2008. Accuracy and clinical implications of seven 25-hydroxyvitamin D methods compared with liquid chromatography-tandem mass spectrometry as a reference, *Ann Clin Biochem*, 45: 153–159.
31. Chen, H., L.F. McCoy, R.L. Schleicher, C.M. Pfeiffer, 2008. Measurement of 25-hydroxyvitamin D₃ (25OHD₃) and 25-hydroxyvitamin D₂ (25OHD₂) in human serum using liquid chromatography-tandem mass spectrometry and its comparison to a radioimmunoassay method, *Clin Chim Acta*, 391: 6–12.
32. Saenger, A.K., T.J. Laha, D.E. Bremner, S.M. Sadrzadeh, 2006. Quantification of serum 25-hydroxyvitamin D(2) and D(3) using HPLC-tandem mass spectrometry and examination of reference intervals for diagnosis of vitamin D deficiency, *Am J Clin Pathol*, 125: 914–920.
33. Priego, C.F., J. Ruiz Jimenez, J.M. Mata Granados, M.D. Luque de Castro, 2007. Identification and determination of fat-soluble vitamins and metabolites in human serum by liquid chromatography/triple quadrupole mass spectrometry with multiple reaction monitoring, *Rapid Commun Mass Spectrom*, 21: 1745–1754.
34. Maunsell, Z., D.J. Wright, S.J. Rainbow, 2005. Routine isotope-dilution liquid chromatography-tandem mass spectrometry assay for simultaneous measurement of the 25-hydroxy metabolites of vitamins D₂ and D₃, *Clin Chem*, 51: 1683–1690.
35. Tsugawa, N., Y. Suhara, M. Kamao, T. Okano, 2005. Determination of 25-hydroxyvitamin D in human plasma using high-performance liquid chromatography-tandem mass spectrometry, *Anal Chem*, 77: 3001–3007.
36. Weiskopf, A.S., P. Vouros, J. Cunniff, E. Binderup, F. Bjorkling, L. Binderup, M.C. White, G.H. Posner, 2001. Examination of structurally selective derivatization of vitamin D(3) analogues by electrospray mass spectrometry, *J Mass Spectrom*, 36: 71–78.

37. Albrecht, A., I. Vovk, 2012. Applicability of analytical and preparative monolithic columns to the separation and isolation of major whey proteins, *Journal of chromatography A*, 1227, 210-218.
38. Deconinck, E., S. Crevits, P. Baten, J. De beer, 2011. A validated UHPLC method for quantification of folic acid in pharmaceutical preparation, *J. Pharm. Biomed Anal*, 54(5): 995-1000.
39. Irena, B., S. Magiera, 2011. Development and Validation of UHPLC method for the determination of flavonoids in red wine, *Journal of AOAC international*, 94(3): 786-794.
40. More, M.K., K.B. Chandrashekhar, Vyas. S, 2011. Degradation studies of triprolidine: isolation, characterization of oxidative degradation products and development of validated stability indicating UHPLC method, *Journal of liquid chromatography and related technologies*, 34(8), 652-669.
41. Kanenik, Z., F. hadacek, M. Marekova, D. Ulanova, J. Kopecky, V. Chobot, K. plhackova, J. Olsovska, 2010. Ultra high pressure liquid chromatography fingerprinting method for chemical screening of metabolites in cultivation broth, *Journal of chromatography A*, 1217(51), 8016-8025.
42. Christopher, C., B. Bailey, A.C. Worth, UHPLC analysis of underivatized amino acids, 2012.
43. Perrin, C., L. Meyer, 2003, *J.Am.Oil.Chem*, 80(2): 115-118.
44. Wolfender, J.L., G. Marti, 2010. Advances in techniques for profiling crude extracts and for the rapid identification of natural products: dereplication, quality control and metabolomics, *Current organic chemistry*, 14(16): 1808-1832.
45. Padmaja, P., R. Wilhard, 2012. Increased throughput and reduced solvent consumption for the determination of isoflavones by UHPLC.
46. Liu, S., J. Xing, Z. Zhang, 2012. UHPLC TQMS inhibitors fishing assay. A novel method for the simultaneously screening of xanthine oxidase inhibitor and superoxide anion scavenger in a single analysis, *Analytica chimica acta*, 715: 64-70.
47. Elia, G., G. Davy, J.L. Wolfender, et.al, 2009. Metabolite profiling of plant extracts by UHPLC at elevated temperatures coupled to TOF-MS, *Journal of chromatography A*, 1216(30): 5660-5668.
48. Kanthithilini, M.K., A.D. Villiers, 2011. Recent development in HPLC separation of phenolic compounds, *Journal of separation science*, 34(8): 854-876.
49. Wolfender, J.L., P.J. Eugster, N. Bohni, M. Ceundet, 2011. Advanced method for natural product drug discovery in the field of nutraceuticals, *Chimia (Aarau)*, 65(6): 400-406.